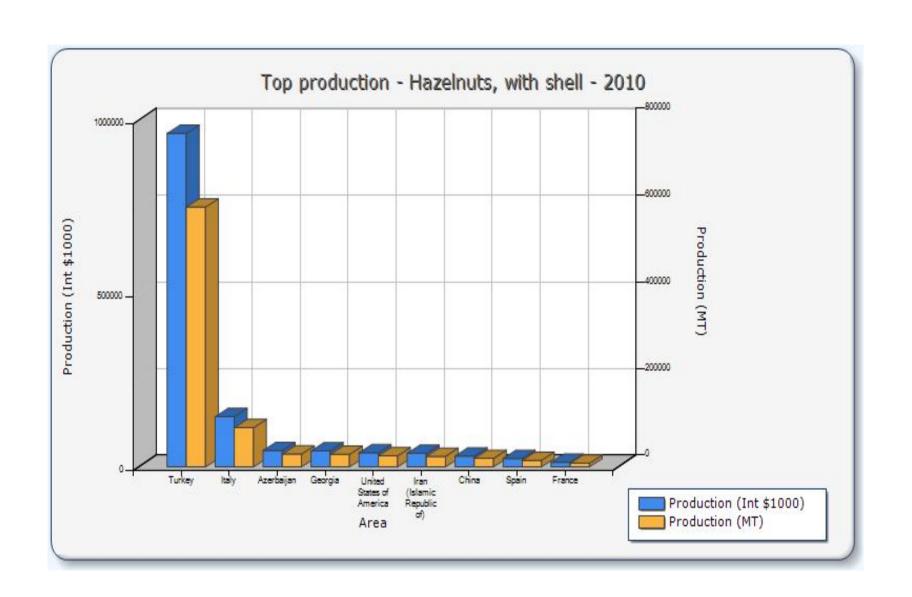


Corylus avellana



Rank	Area	Production (Int \$1000)	Flag	Production (MT)
1	Turkey	961744	*	600000
2	Italy	144694	*	90270
3	Azerbaijan	47212	*	29454
4	Georgia	46163	*	28800
5	United States of America	40715	*	25401
6	Iran (Islamic Republic of)	38950	*	24300
7	China	31256	*	19500
8	Spain	24203	*	15100
9	France	13284	*	8288
10	Croatia	5342	*	3333
11	Kyrgyzstan	5289	*	3300
12	Poland	4007	*	2500
13	Greece	2083	*	1300
14	Belarus	1923	*	1200
15	Tajikistan	1763	*	1100
15	Uzbekistan	1763	*	1100
17	Russian Federation	1282	*	800
18	Armenia	801	*	500
19	Mongolia	625	*	390
20	Republic of Moldova	480	*	300



سطح زیرکشت، میزان تولید و عملکرد محصولات باغی (دایمی) کشور به تفکیک محصول در سال ۱۳۸۷

جدول شماره ۲–۱ ((واحد: هكتار))

نام محصول	سطح بارور			سطح غيربارور		جمع سطح بارور و غیربارور			
نام محصول	آبی	ديم	جمع	آبی	ديم	جمع	آبی	ديم	جمع
سته	۲۷۹1۷ ٦,۷	71,5	479197	1,50010	٩,٥٥	01907	£41.47,4	٧٧,٣	27110+
دام	19-01,1	0411-,4	127771,4	71701,1	7,14871	TATT ,9	11-4-7,0	YE097,T	140298,4
ردو	10+017,7	3,7070	100718,7	0,79730	۲,۱۱۵۳	049+0	۲٠٤٩٠٥,٨	۸۷٦٣,٩	T17779,V
ندق	14.04,8	۹۷۸۸,۹	۲۲۸٤٧,٣	15444	۲۲۸	74557	15044,4	1.708,9	10191,1

سطح زیرکشت، میزان تولید و عملکرد محصولات باغی (دایمی) کشور به تفکیک محصول در سال ۱۳۸۷

((واحد: تن - كيلوگرم))

ادامه جدول شماره ۲-۱

عملكرد		توليد			1
ديم	أبى	جمع	ر ديم جمع	أبى	نام محصول
101,9	0.4,1	197779	٣	197777	پسته
٤١٣,١	1100,7	177779	17747	1+ TAAT	بادام
TTE1,9	78.7,1	779171	14-44	777127	گردو
1144,7	1272,9	T-72T	11777	147.4	فندق

GROUP	SPECIES	AREA OF DISTRIBUTION		
Avellana group (shrub)	Corylus avellana L. Corylus maxima Mill. Corylus Americana Marsh. Corylus heterophylla Fish.	Europe, Anatolia, Caucasus, Urals Anatolia, Eastern Europe North America Manchuria, Korea, Japan		
Cornuta group (shrub)	Corylus cornuta Marsh. Corylus californica Rose Corylus sieboldiana Blume	North America North America Mongolia, Manchuria, Korea, Japan		
Colurna group (tree)	Corylus colurna L. Corylus chinensis Franchet Corylus jacquemontii Decaisne Corylus papyracea Hickel Corylus ferox Wallich	Eastern Europe, Anatolia, Caucasus to the Himalayas Central China Afghanistan, northern India China Intermediate zone of the Himalayas central China, Nepal, Sikkim		

Type of organ	Catkin	Inflorescences (female)	Vegetative buds
Number of hours at T < 45° F	< 100 to 365-480	480-600 to 1170-1255	600-680 to 1170-1255

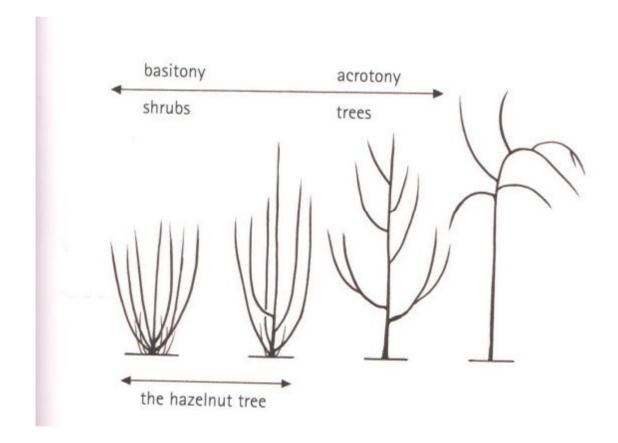
Frost damage on a shoot





Basitonic growth throughout the tree

The hazelnut tree is a shrub



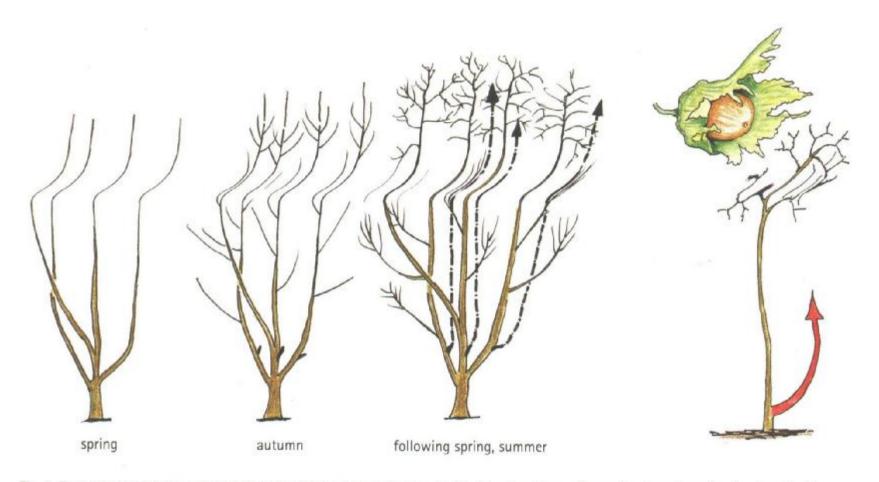
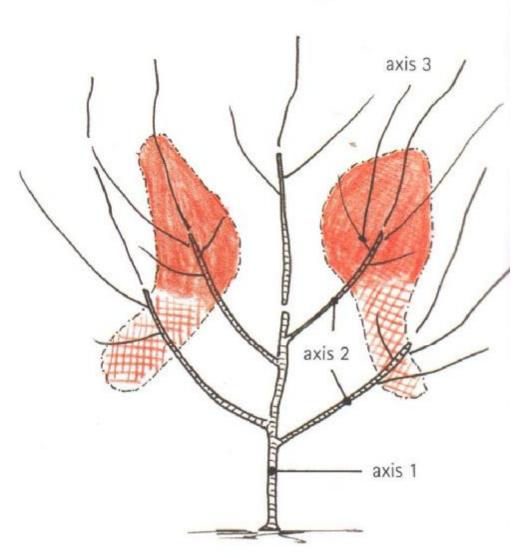


Fig. 2. The process of epicormic branch formation on hazelnut trees: basitonic reiterations often arise when there is abundant fruiting.



The areas of first fruiting on a tree that was not cut back at planting







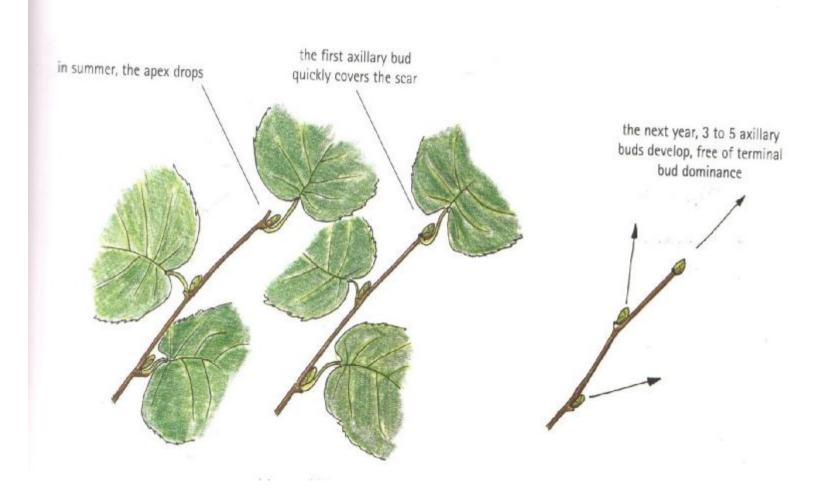


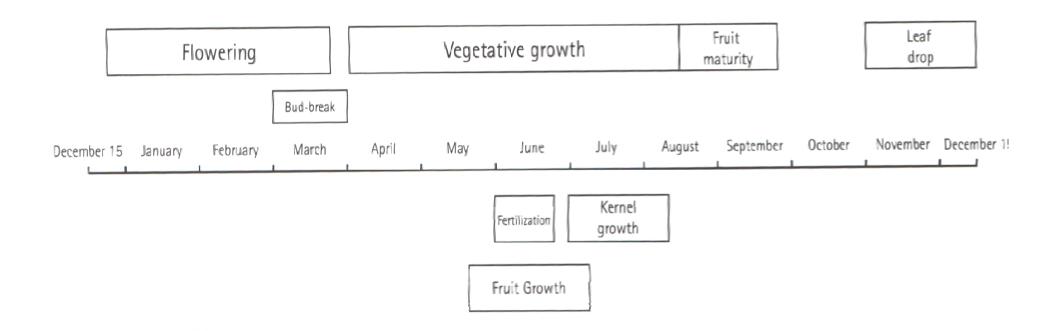




















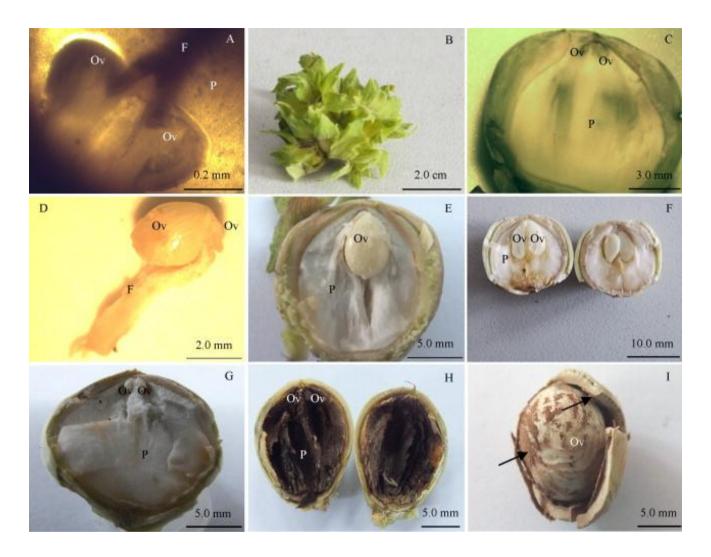










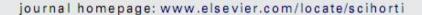


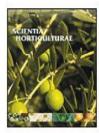
Ovule filling and blank fruit formation. (A) Two ovules formed in an ovary, 22 May; (B) fruit cluster, 23 June; (C) fruit at early development stage, with one developed and one undeveloped ovule. Most space inside the fruit shell was filled with parenchyma, 2 July; (D) two ovules attached with funiculus, one ovule developed (left) and the other one (right) cease development, 12 July; (E) rapid expanding ovule, only one ovule developed in one kernel, 2 August; (F) double-kernel fruit, two well developed ovules, 2 August; (G) blank fruit. Two ovules ceased development and most of the inside shell was filled with parenchyma, 2 August; (H) blank fruit at harvest. Two undeveloped ovules were left inside the shell, 22 August; (I) normal filled kernel at harvest. Blown parenchyma residue was shown by arrows, 22 August. P: parenchyma; F: funiculus; Ov: ovule.



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The relationship between reproductive growth and blank fruit formation in *Corylus heterophylla* Fisch

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ARTICLE INFO

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Keywords: Hazelnut Blank fruit Embryo abortion

ABSTRACT

The reason leading to the high blank fruit ratio of *Corylus heterophylla* Fisch was elucidated by investigating pollen compatibility, ovule and embryo development. It was showed that the female flowers bloomed in the middle of April and the ovary did not develop until 1 month later. In late May, two ovules were found in an ovary. On 22 June, the ovule wall differentiated into integument, and nucellus endosperm were clearly observed in the ovules, suggesting the starting of fertilization event. Globular, heart, torpedo and cotyledon embryo developed step by step from 28 June to 12 July. The ovule grew rapidly in filled fruit since 12 July. Ovule in blank fruit ceased growth from 12 July although the full embryo with cotyledon could be observed. The blank fruit could be distinguished from the filled one for its undeveloped ovule and large amount of parenchyma. There was no significant difference in ovary size between the filled and empty nuts, but the weight of blank nut was only about one half of the filled one. It is concluded that formation of blank fruit of *C. heterophylla* Fisch is closely related to embryo abortion, but not incompatibility between the pollen and stigma.



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An investigation of the relationship between reproductive growth and yield loss in hazelnut

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 Received 7 December 2005; received in revised form 13 October 2006; accepted 21 February 2007

Abstract

This study was carried out in Samsun during a 2-year period to examine the relationship between reproductive and yield loses in the 'Tombul' and 'Palaz' hazelnut cultivars. In hazelnuts, male and female flowering occur in winter after the breaking of inflorescence dormancy. In the present study, growth of the ovary of the hazelnut started in April and continued until mid-June. At the time of flowering the ovary did not form. The ovule growth showed a rapid increase at the end of June. Change in the diameter of the ovary and ovule with time showed a simple sigmoid growth curve. Fertilization occurred during the period between mid-May and the beginning of June, namely, 3.5–5 months after pollination. At this time, the diameter of the nut was 9.54 mm. Twin kernel was not observed. The ratio of double kernels was close to zero. The time period from fertilization to barvest was 89 days in 1997 and 96 days in 1998 for Tombul cultivar. For the Palaz, this period was 84 days in 1997 and 86 days in 1998. The rate of pistillate flower clusters which dropped in April–May was more than those dropped in June–August.

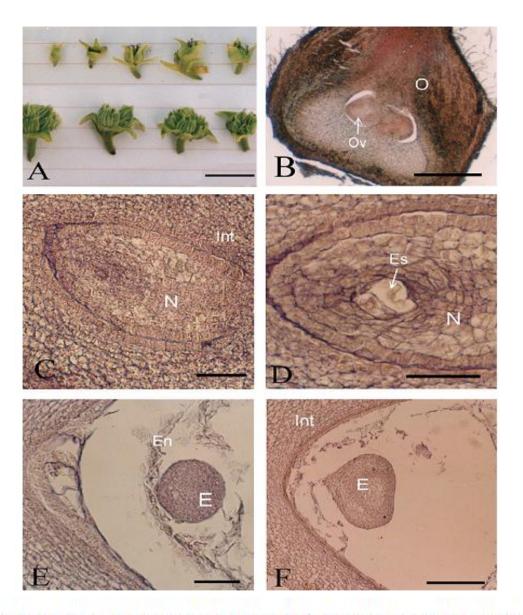


Fig. 1. Early stage ovary, ovule and embryo development. (A) Macrophotograph of the clusters from female flowers with non-developed ovary (above), and developing some of the ovaries (below), 30 April. (B) Micrograph of the ovary longitudinal section. The two ovules are visible on the placenta, 7 May. (C) Nucellus forming, 21 May. (D) Nucellus with embryo sacs, 28 May. (E) Globular embryo and cellular endosperm, 4 June. (F) Heart-shaped embryo, 13 June. Bars A, 1 cm; B, 0.5 mm; C-E, 0.05 mm; F, 0.2 mm (Es: embryo sac; O: ovary; Ov: ovule; N: nucellus; Int: integument; E: embryo; En: endosperm).

According to our result, difference in the percentage of the pistillate flower cluster dropping between years in the same cultivar was evident. One of the reasons as to the early clusters dropping was lack of pollination. Thompson (1967) clearly indicated that growth of the ovary was dependent upon pollination. If growth is arrested in all of the flowers in a cluster, the cluster withers and falls in late April or May. Both Thompson (1967) and Beyhan and Odabaş (1996) reported that some ovaries could not grow more than 0.5 mm and these pistillate flowers dropped in April and May. On the other hand, while pollination is necessary to initiate ovary development; only a small percentage of the pollinated pistils develop into a full sized nut.

In the present study, in May, it was determined that the average number of functional ovaries in one cluster was 40-60% for the Tombul cultivar and 40-55% for the Palaz cultivar. These results showed a similarity with the findings of Beyhan (1995) in which 45-60% of the ovaries could not develop further. Germain et al. (1978) and Germain (1994) reported that, shortly after leafing, up to 70% of the flowers can stop growing.

When all the flowers in a cluster are affected by this phenomenon, the cluster drops by the end of April or in May. However, Dimoulas (1979) reported that, in general, these drops were not caused by a lack of pollination or by competition between flowers within the same inflorescence but were closely related to apical dominance which occurs along the 1-year-old shoots and the peduncles of catkins.

The ovaries and ovules of pollinated pistillate flowers continue to develop until fertilization. But not all pistillate flowers are fertilized. The diameter of the ovary was found to be 9.54 mm during this period in our study which is a critical phase for the hazelnut. As Silva et al. (1996) stated, ovules will not develop in non-fertilized ovaries and blank fruits form. On the other hand, even if fertilization occurs, the ovule and development of the embryo may cease at various times during the period from fertilization to harvest. The clusters with ovaries of both type drop. If even one ovary is fertilized and it develops into a cluster, that cluster will not drop.

Mehlenbacher et al. (1993) reported that nut and kernel defects are serious problems for the hazelnut. These include blanks, brown stain disorder, doubles, moldy kernels, kernels with black tips, shriveled kernels and poorly filled nuts. Brown stain, a poorly understood disorder that leads to kernel abortion, appears sporadically in Oregon and can result in severe crop loss. Poorly filled nuts and shriveled kernels are defects in which the kernel is smaller than in size. Small kernels are most common when the crop load is heavy or trees are stressed during the period of rapid kernel growth.

Lagerstedt (1977, 1985) reported that cluster droppings were caused by the genetic constitution of the cultivar, alternate bearing habit, pollen source, sexual incompatibility, cultural practices (nutritional deficiencies, lack of irrigation, disease and insect pests), and environmental conditions. On the other hand, Beyhan and Marangoz (1999) reported that the fruit cluster drops which occurred in June were mostly due to nutritional deficiencies while the cluster drops which occurred in July or August were caused by insufficient soil moisture and nutritional deficiencies.

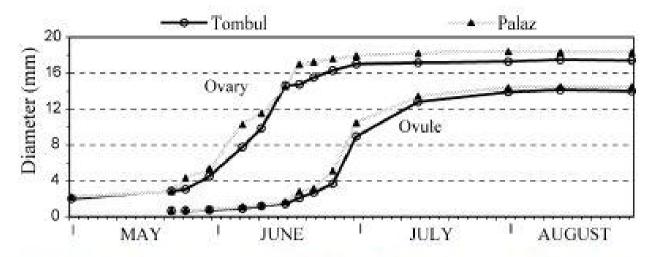


Fig. 3. Growth curves of ovary and ovule diameter of equivalent sphere for Tombul and Palaz hazelnut in 1997.

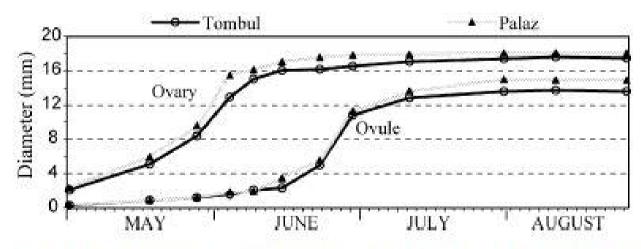
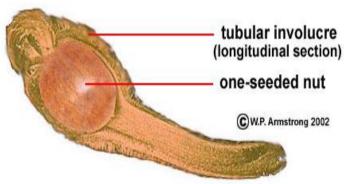
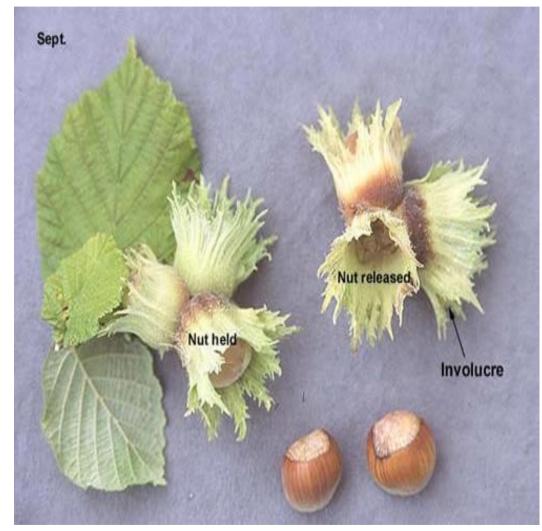


Fig. 4. Growth curves of ovary and ovule diameter of equivalent sphere for Tombul and Palaz hazelnut in 1998.





Beaked Filbert or Hazelnut (Corylus cornuta)









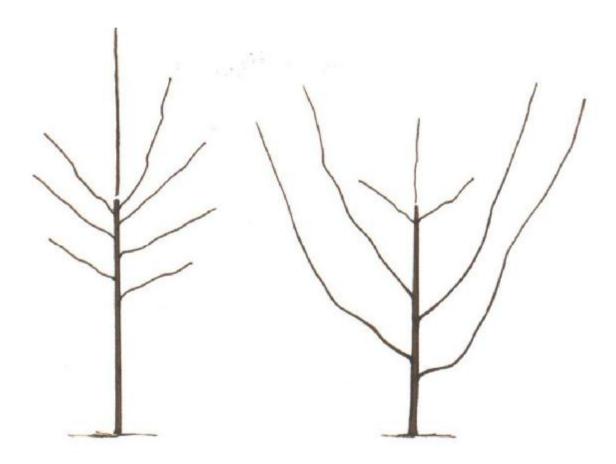






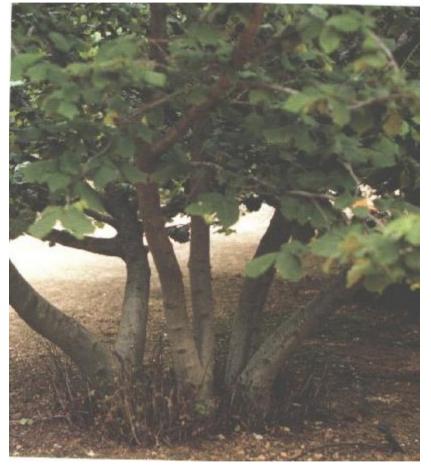


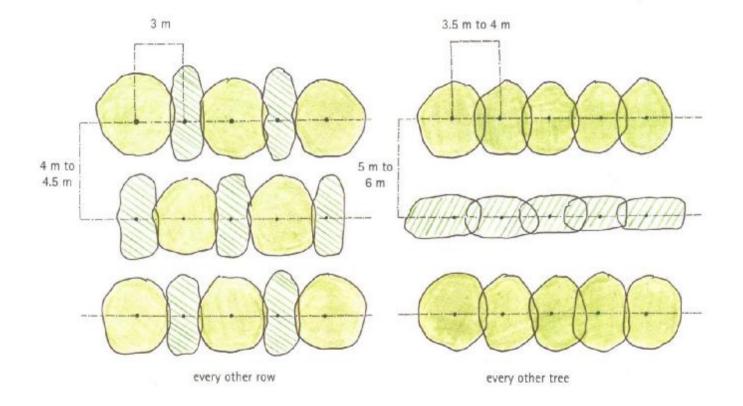




Two behaviors of first year hazelnut trees

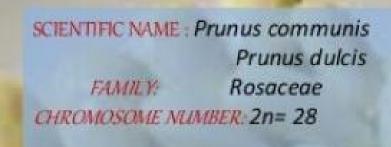






Almond





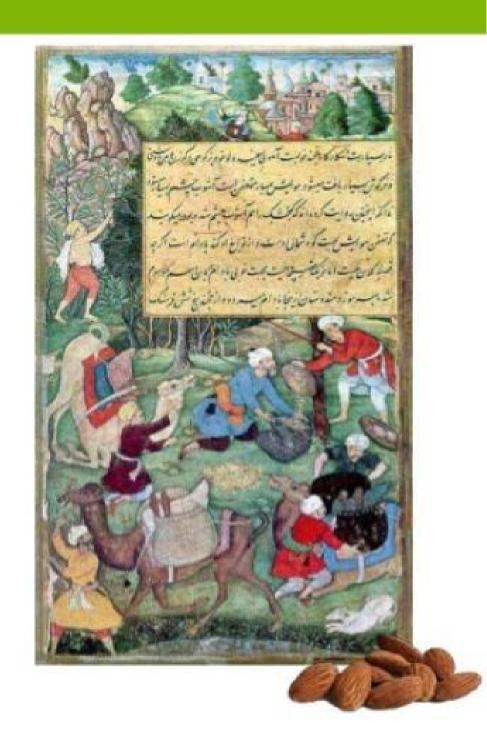
ORIGIN AND DISTRIBUTION:

- >Originated from south west and central Asia
- Major Growing Countries: California, Spain, Italy, Greece, Turkey, India
- California is the leading producer of almond in world with 489,879,76 ton
- India's production: 1,179,34 ton
- > major states in India:
- *Jammu and Kashmir (Almond is the state tree of Jammu and Kashmir)
- sub tropical varieties of almond is grown in North Indian conditions.

- Prehistory
 - Wild almonds emerge on mountains separating China from Kazakhstan, Afghanistan and Iran
- 4,000 B.C.
 - Almonds cultivated in Central Asia and eastern Mediterranean
- 1,352 B.C.
 - King Tut takes handful of almonds to his grave
- 350 B.C.
 - Alexander the Great's armies bring almonds to Greece



- 1500's
 - Harvest in the Fergana Valley Uzbekistan



- 1700's
 - Brought to California from Spain by Franciscans
- 1840's
 - California growers experiment with almonds
- 1880's
 - California almond crop tops one million Lbs.
- 1986
 - "A Can A Week Is All We Ask" ad begins
- 2002
 - California almond crop tops one billion Lbs.
- · 2014 +
 - California crop expected to top two billion Lbs.





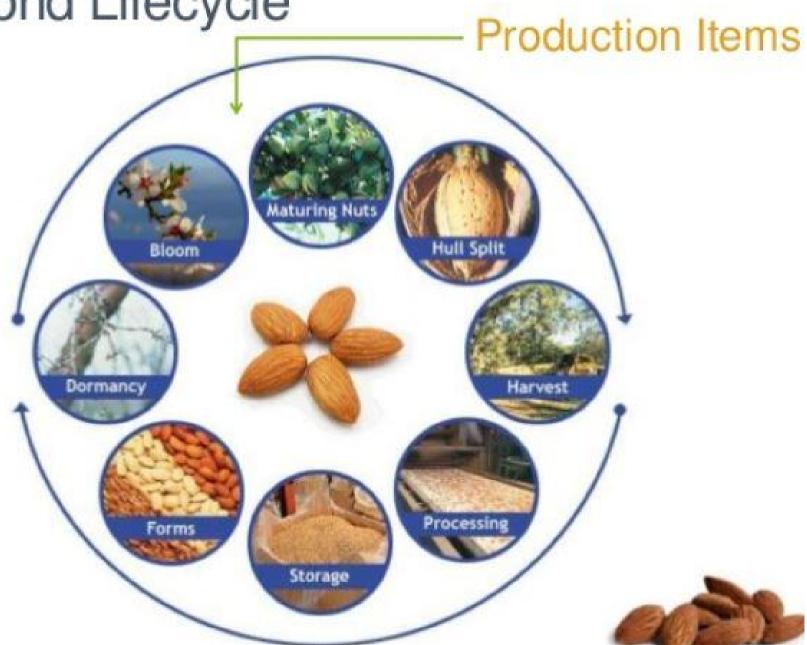
Blossoming Almond Tree - Vincent Van Gogh (1890)



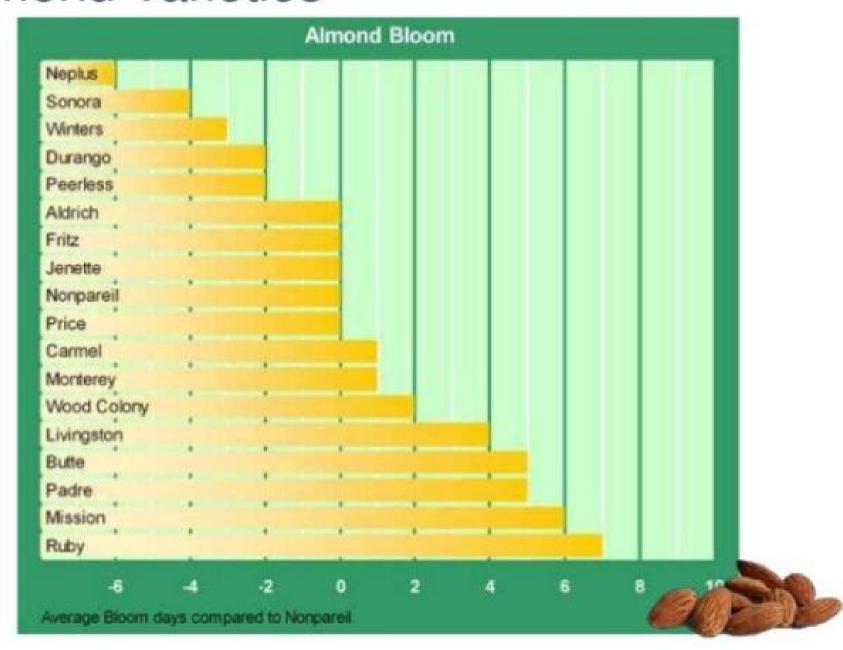
Blossoming Almond Tree - iPhone Case (2013)



Almond Lifecycle



Almond Varieties



MAJOR CALIFORNIA ALMOND VARIETIES

	CHARACTERISTICS CLASSIFICATION						
variety		TAMBARIA	SHORT A PLUMP/POUPED	CALIFOGRAM	Messon The Series	Monthlyness	PASSIELL HAND MESS.
NONPAREIL (NP)	SHELL Soft shell, ight coer, high sufureopening NUT Medirm, flat shape, smooth surface	•				•	
CARMEL (CR)	SHELL Bott shell, good shell integrity, fair suture opening NUT Medium, narrow shape, slightly wrinked surface	0		0			
BUTTE (BT)	SHELL Sami-hard shell, light color, smooth surface, low auture opening NUT Small, short plump shape, winkled surface		•	0	0		
PADRE (PD)	SHELL Hard shell; good shell htegrity, no suture opening NUT Small, short wife shape, winked surface		•	•	•		
MISSION (MI)	SHELL Hard shell good shell htegrity, no suture opening NUT Small, short wife shape, dark brown, deeply		0		0		

	CHAR	AR ACTERSTICS			DASEPIGNICA			
		SERVICE S. ILAST	SHORT A PLAINWANDUND	CALFFORNIA	No.	Month decid	Man State	
MONTEREY (MT)	SHELL Hard shelt, smooth surface, low suture opening NUT Large, long narrow shape, deeply winked surface	•		0				
SONORA (SN)	SHELL Soft shell, dark brown colourough surface, high surfureopening NUT Large, long narrow shape, light coor, smooth surface	•		•				
FRITZ (FR)	SHELL Bem-hard shell, good shell integrity, low acture opening NUT Bmail medium plump shape, fairty winkled surface		•	•	•			
PRICE (PR)	SHELL Soft shell, dark brown colourough surface, high sutureopening NUT Small, short narrow shape, fairly winked surface	•		•				
PEERLESS (PL)	SHELL Hard shell, good shell integrity, smooth surface, no suture opening NUT Medium, wide shape, fairly winked surface	•		0			0	

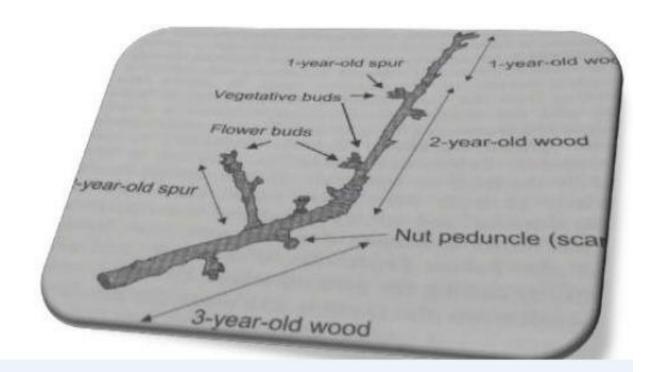


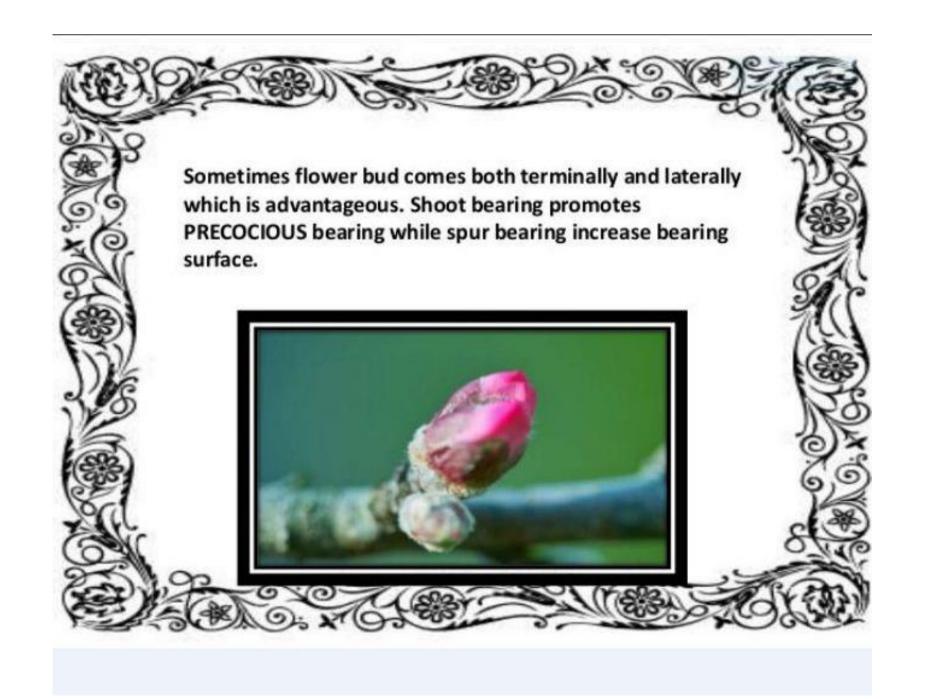
FACTOR	PARAMETERS	COMMON TERMINOLOGY					
VARIETY	Shape, color, skift texture or smoothness, blanchability	Nonparei J Carme J Butte, Padre, Mission, Monterey, Sonora, Fritz, Paerless, Price					
SIZE"	Count range of whoe almond kernes per ounce (2835 grams)	18/20, 20/22 23/25, 25/27, 27/30,30/32, 32/34, 34/36,36/40, or customer-specified range					
GRADE	Disamila_rdoubles, chipped and scratched kemels, foreign material, split and broken kemels, other defects and serious damage	Fancy, Extra No. 1, No.1 (Supreme), Select Sheller Run, Standard Shell Run, No. 1 Whole and Broken, No. 1 Pieces					
Shelihardness, shell stegrity, sutureopening, kerne iquality, drack out		Market specific, depending on how in-shell will be ultimately soid to consumers, for example: Traditionals sold in the shell—earth-or hard she specifishel, cracked within mechanish nut crac Shacks sold in the shell—soft shell with greater siture opening to allow seasonings topermeate the shell. Hand Gracks sold as kernels—soft shell preferred to allow manual cracking.					

"notividualishoe lianet stamayyary horiyaar to yar asa say tof variatina inwaathat powing conditions and poduction yalds there are said stilly present staathat be limited in someyabre.



- *Botanically Almond is a Drupe
- •Edible part is nut
- Inflorescence come laterally on current season growth which bear fruit in following season.







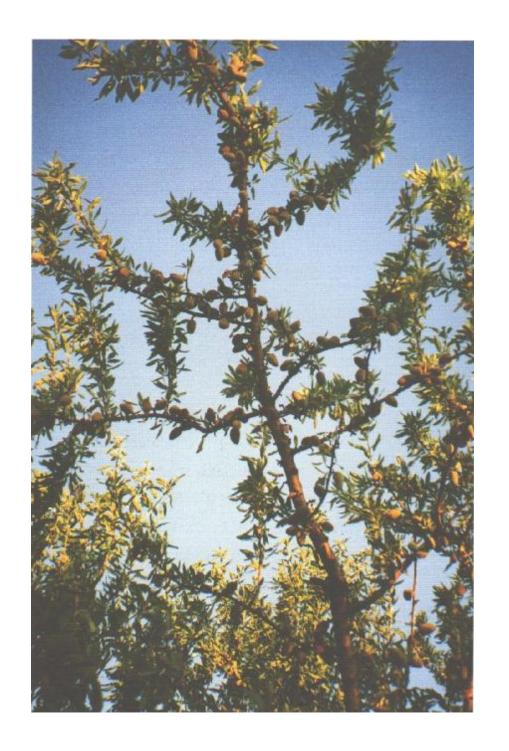


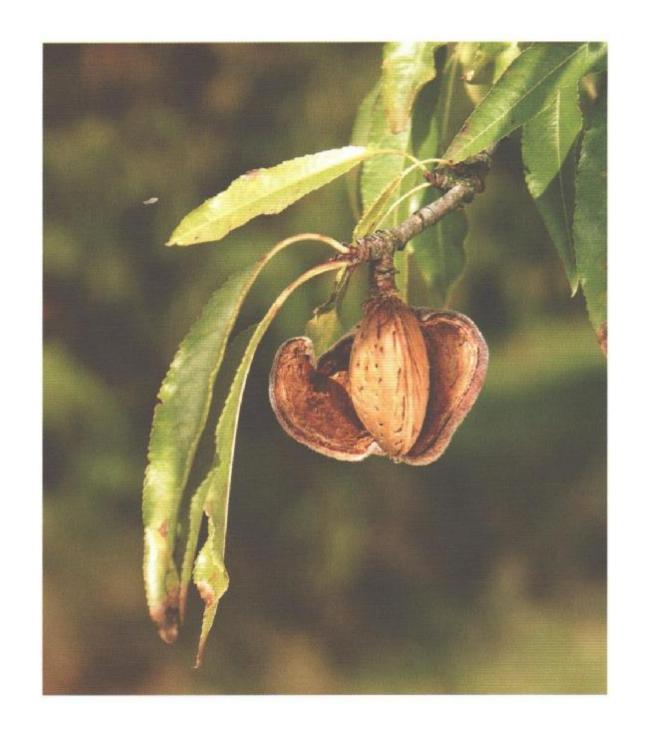


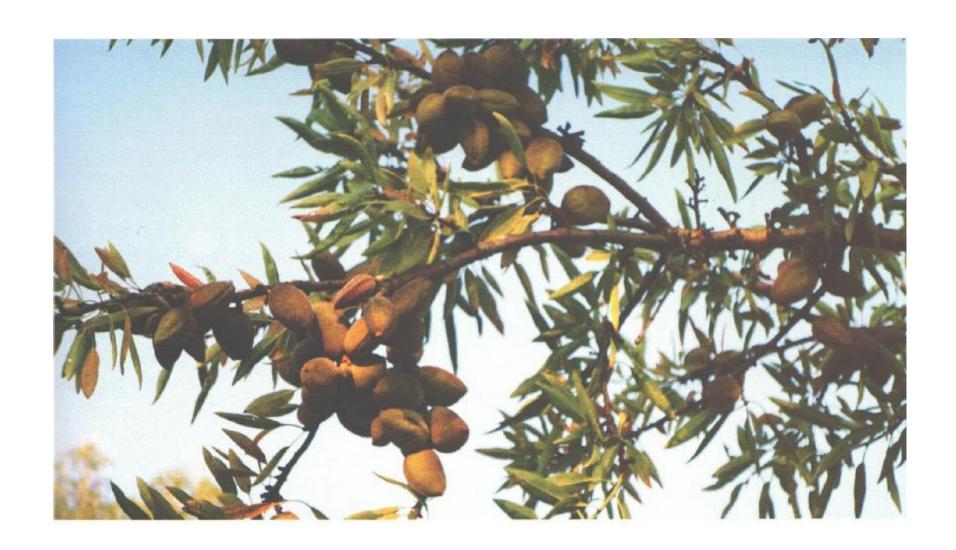
INFLORESCENCE OF TEMPERATE ALMOND

INFLORESCENCE OF SUB-TROPICAL ALMOND









Bud Break / Bloom

February and March



Bloom



Bloom Sprays



Pollination

- Single most important yield factor
 - Hive management critical
 - 2 Hives per Acre at bloom
 - \$150 -\$200/Hive
- Colony Collapse Disorder
 - Varroa mites
 - Neonicotinoids
 - Habitat loss
 - Virus
 - American foulbrood disease





Pollination



Maturing Nuts

April - June





Hull Split

July - August



Harvest

- August October
- Shaking
- Raking / Sweeping Dry on orchard floor
- Pickup
- Hulling / Shelling
- Processing



Harvesting:

HARVESTING TIME: Early August to Late September

HARVESTING INDICES:

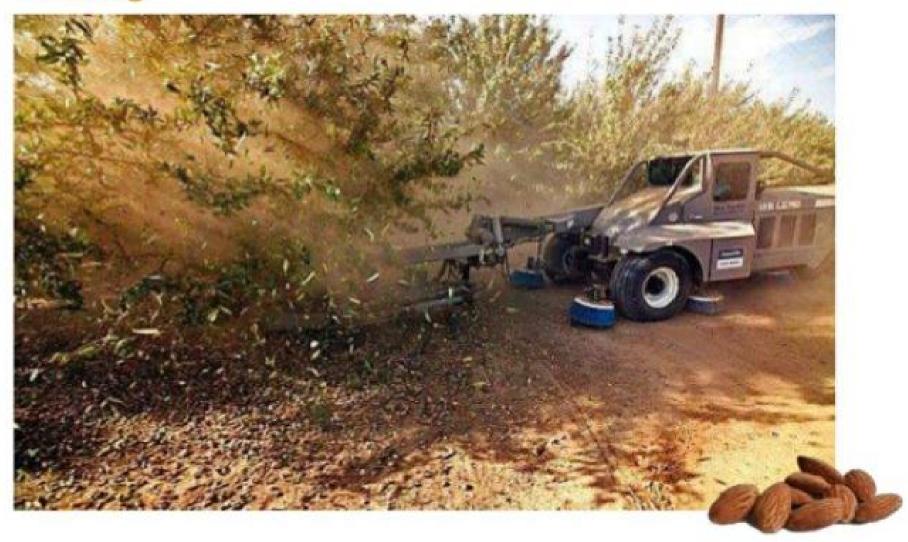
- •Hull split of 95% of nuts
- Ripened nuts should be prevented from bird damage

HARVESTING METHOD:

Almond tress are knocked and falling nuts are collected in net

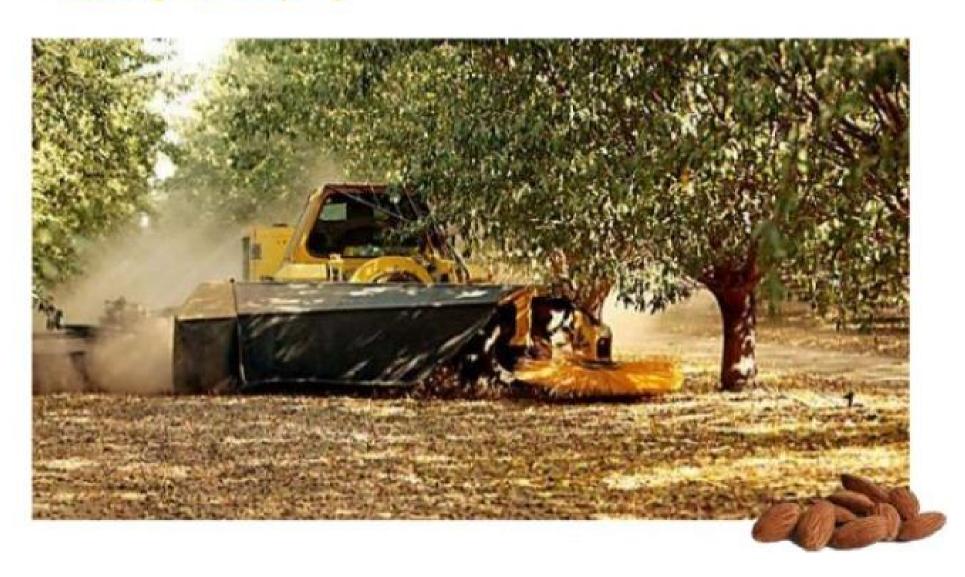
Almond Harvest

Shaking



Harvest

Raking / Sweeping



Harvest

Hulling





Harvest

Hulling



Processing - Products







INFLORESCENCE OF TEMPERATE ALMOND

INFLORESCENCE OF SUB-TROPICAL ALMOND





- Before going for storage nuts should be treated at -17.8 *c for 48 hours to destroy the eggs and pest of previously applied naval orange pest
- Almond can be stored for * months under room temperature
- It can also be stored for more years if kept at 0 to 7 *C



Water Management

- Central Valley is a Mediterranean Climate
 - · Cool, wet winters and warm, dry summers
- Limited Water Supplies
 - Drought conditions
 - Regulatory issues
- Use roughly 3.5 4.0 Acre Feet / Acre
 - Drip irrigation is method of choice
 - Soil amendments, fertilizers through drip



Irrigation

- Methods of Irrigation
 - Flood
 - Drip
 - Micro



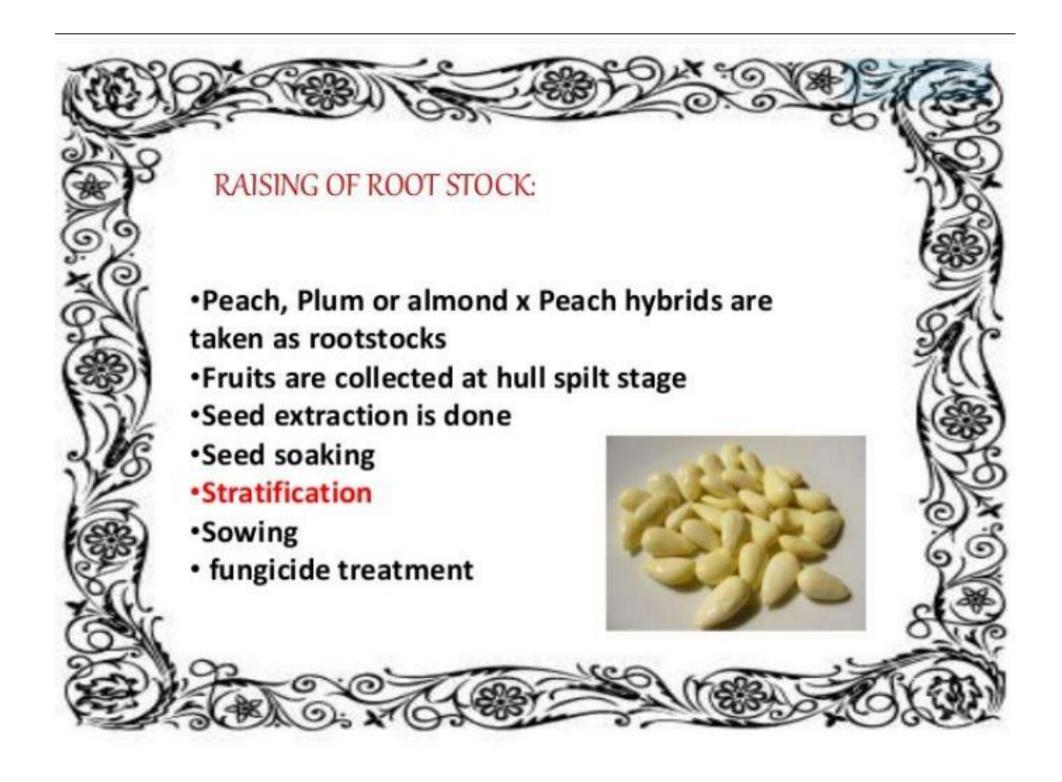
PROPAGATION:

TIME: Late spring, root stock grown in

autum

METHOD: T-Budding





Cultivation

Planting

- Almond plants are planted in 1m x 1mx 1m sized pits at a distance of
 4.5 m x 4.5m in January in square system.
- •Most of the almond varieties are self-unfruitful. Therefore to plant two or more cross-compatible varieties which flower at the same time in alternate rows are essential.

Training and pruning

- Almond plants are trained according to modified leader system.
- •It bears mostly on spurs which live for five years. Thus in young trees, only diseased, dry wood and the branches which are interfering with each other should be removed.
- Water sprouts arising on the stem should also be removed as early as possible.
- •In old bearing trees, pruning is done to remove one-fifth of the growth every year.

FERTILIZATION AND IRRIGATION:

- *As mesophytic origin crop it required less water.
- Irrigation should be done thoroughly but not so frequently as it is suseptible to water logging conditon
- Irrigation should be with holded prior to harvest as the result ing water stress promotes abscission layer
- It mainly required Nitrogen application.
- Almond removes 45kg of actual N from soil
- *Leaf analysis should be done during summer to identify deficiencies
- Under high yield cropping systems pottasium and zinc may be applied as potassium easilyfixed by clay become unavailable for root uptake
- *Zinc or foliar spray of zinc sulphite in autumn helps to remove remaining leaves on tree as they go into winter dormancy